# Antibiotic Resistance Profile of *Escherichia coli* Isolated from Stool Samples of COVID-19 Patients

# Sarah Maher Abou Alfa, Wafaa Mohammed Kamel Bakr, Mohamed Anwar Mahgoub\*

Department of Microbiology – High Institute of Public Health, Alexandria University, Egypt

### **Abstract**

Background: Antimicrobial resistance (AMR) poses a serious worldwide threat besides causing several concerns for the health system. E. coli has evolved to be capable of acquiring antibiotic resistance characteristics from other bacteria in its environment and of being easily transferred via fecal-oral route. However, this concern has not received enough attention during the last COVID-19 pandemic. Aim: To evaluate the antimicrobial profile of fecal E. coli isolated from COVID-19 patients. Subjects and methods: This study was conducted on 180 patients divided into two groups: the group of COVID-19 patients and the group of non-COVID-19 patients (90 each). Fecal samples were subjected to microbiology procedures for isolation and identification of E. coli strains. The identified strains were subjected to antimicrobial susceptibility testing using Kirby-Beuer test (disc diffusion method) and interpreted according to Clinical and Laboratory Standards Institute (CLSI) guidelines. Results: The study found that a substantial proportion (82.2%) of patients with COVID-19 received azithromycin. Furthermore, the COVID-19 group demonstrated a significantly higher resistance to ertapenem compared to the non-COVID-19 group (8.9% versus o%). Notably, E. coli isolates from COVID-19 patients exhibited a considerably higher multi-drug resistance (MDR) profile than those from non-COVID-19 patients (55.3% versus 44.7%). Conclusion: E. coli isolated from fecal samples of COVID-19 patients showed resistance to many tested antibiotics. Although, they showed significant resistance to ertapenem, the most efficient drugs against E. coli were the carbapenems. MDR E. coli was observed in both groups signifying community-acquired resistance. However, significant MDR pattern was detected among COVID-19 patients.

**Keywords:** Pandemic; COVID-19 patient microbiota; Fecal *E.coli*; Multidrug resistance; Gramnegative bacteria.

#### Introduction

Antimicrobial resistance (AMR) is a serious global health concern, with predictions suggesting it could lead to 10 million deaths by 2050. The prevalence of AMR varies significantly across different regions. High-income countries (HICs) also face significant AMR challenges. For example, the United States reports approximately 2.8 million cases of antibiotic-resistant infections annually. In contrast, lower-middle-income

countries (LMICs) struggle to accurately assess the extent of AMR due to limited data. Data on AMR is unavailable for around 42.6% of African countries.<sup>(3)</sup>

During the COVID-19 pandemic, the widespread use of antibiotics, including drugs like azithromycin, both for treating bacterial infections and as an immune system modulator, was associated with an increase in antibiotic resistance. Systematic reviews revealed a low occurrence of bacterial coinfections and

superinfections in COVID-19 patients, despite the excessive use of antibiotics. (4,5) In many instances, the decision to treat COVID-19 patients with antibiotics for bacterial infections lacked solid evidence. These decisions were often based on the severity of the patient's condition or the clinician's prior experience, rather than on established criteria for co-infections or superinfections. (6)

Escherichia coli (E. coli), extensively studied worldwide, serves as a model organism for understanding antibiotic resistance. Its ability to spread resistance genes to other bacteria and contribute to the rise of multidrugresistant (MDR) infections is a major concern. (7,8) This is facilitated by its easy transmission between humans and from animals to humans through the fecal-oral route, making it a significant driver of the global antibiotic resistance crisis. (9)

The rapid bacterial evolution under selective antibiotic pressure is one of the most significant causes that have contributed to the present AMR issue, since a persistent contact between bacteria and any received antibiotic is considered as a vital component for the MDR strains increase. (10)

Regrettably, the global response to this catastrophe has been insufficient despite the global rise in infections brought on by MDR bacteria. Individuals continue to overuse antibiotics and have even intensified this use over time. Analyzing global antibiotic sales statistics, it was shown that, from 2000 to 2015, the rate of antibiotic consumption worldwide grew by 39%, from daily doses/1000 individuals of 11.3 per day to 15.7. (10,11)

According to Klein et al., (11) LMIC account for the majority of the world's use of antibiotics and also have the greatest rates of infections caused by MDR

bacteria. Usage of last-resort antibiotics like colistin and carbapenems is also increasing consistently with the emergence of resistant *E. coli* to these medications. (12)

# **Subjects and Methods**

This study investigated the resistance patterns of *E.coli* isolated from stool samples collected from 90 COVID-19 patients and 90 non-COVID-19 patients.

Study Design and Setting: This comparative cross-sectional study was carried out during a period of eighteen month, from February 2021 through September 2022. The patients included in this study were recruited from a private diagnostic laboratory in Alexandria, where they had visited for routine health checkups and diagnostic evaluations.

# Study Population Inclusion criteria:

-Non hospitalized COVID-19 patients with confirmed reverse transcription polymerase chain reaction (RT-PCR) result and / or clinical or radiological findings, who received antibiotic therapy during the duration of their COVID-19 infection.

-Non-COVID- 19 patients who received antibiotic therapy in the last six months before the study.

#### **Exclusion criteria:**

- -COVID-19 patients with associated other diseases that required antibiotic treatment.
- -Patients less than 18 years old.
- -Non COVID-19 patients with history of COVID-19 or in contact with positive cases of COVID-19.

# Sample size

Based on a previous study by Shakya et al.<sup>(13)</sup>, a minimum required sample size of 90 stool samples of COVID-19 patients as well

as 90 stool samples of non-COVID-19 patients achieve 80% power to estimate the proportion difference of antibiotic resistance of E. coli isolated from stool samples between the two groups. Sample size was estimated based on an average antibiotic resistance prevalence rate of 33% based on another published study from healthy subjects. An increase of 20% in antibiotic resistance pattern among COVID-19 patients was anticipated. Calculation was performed by comparing two binomial distributions at 0.05 level of significance. Sample size was calculated using MedCalc software.

# **Collection of samples**

Stool samples were collected in sterile plastic cups and transported to the microbiology laboratory at the High Institute of Public Health, Alexandria

University within 2 hours in an ice box. For each patient enrolled in this study, a comprehensive sheet was filled out, containing information such as sampling date, time of collection, name, age, sex, type and duration of antibiotics intake, and other relevant clinical data. Consecutive samples were collected till reaching the required sample size.

# Isolation and Identification procedures:

All stool samples were directly plated using sterile loop on MacConkey's agar plates. Plates were aerobically incubated overnight at 37C°. Suspected isolated E. coli colonies were subcultured on blood agar plate for confirmation using standard microbiological procedures. (14,15) These procedures included macroscopic, microscopic and biochemical identification as shown in table (1).

Table (1): Biochemical tests for E. coli.						
E. coli	Results					
Triple sugar iron						
Slant/butt	Acidic/Acidic					
H <sub>2</sub> S	Negative					
Gas	Positive					
Indole production	Positive					
MR	Positive					
VP	Negative					
Citrate utilization	Negative					
Urease	Negative					
Motility	Positive					

Macroscopically, they showed bright pink lactose fermenting colonies on macConkey agar plates and non hemolytic non pigmented greyish convex smooth colonies on blood agar plates. *E.coli* appeared as rod shaped Gram negative bacteria with no special arrangement under the microscope.

All the identified *E. coli* isolates were subjected to antimicrobial susceptibility testing on Mueller Hinton agar (MHA) plates using disc diffusion method according to the Clinical Laboratory Standards Institute guidelines (CLSI) as illustrated in table (2). (16)

Table (2): Interpre	tive categories and zo	ne diameters br	eak points for E	interobacterales				
Test/Report	Antimicrobial	Disc content	Zone Diameter Interpretive Criteria (nearest whole mm)					
Group	agent		Susceptible (S)	Intermediate (I)	Resistant (R)			
Penicillins								
Α	Ampicillin	10 µg	≥17	14-16	≤13			
β-Lactam/β-lactam	nase inhibitor combinat	ions						
В	Amoxicillin-clavulana	20/10 μg	≥18	14-17	≤13			
Cephems (parente	eral)							
В	Ceftriaxone	30 μg	≥23	20-22	≤19			
Α	Cefazolin	30 µg	≥23	20-22	≤19			
В	Cefepime	30 μg	≥25	19-24	≤18			
Carbapenems								
В	Ertapenem	10 μg	≥22	19-21	≤18			
Aminoglycosides		,	-					
Α	Gentamycin	10 μg	≥15	13-14	≤12			
Quinolones and flu	uoroquinolones				•			
В	Cirpofloxacin	5 μg	≥26	22-25	≤21			
Folate pathway ar	ntagonists				•			
В	Trimethoprim- sulfamethoxazole	1.25/23.75 μg	≥16	11-15	≤10			
Tetracyclines								
0	Doxycycline	30 μg	≥14	11-13	≤10			
Monobactams								
C	Aztreonam	30 µg	≥21	18-20	≤17			

# **Ethical considerations**

An informed written consent was obtained from all participants after explanation of the purpose of the study. The study was approved by the Ethics Committee at the High Institute of Public Health (HIPH), Alexandria University.

# Statistical analysis of the data

Data were fed to the computer and analyzed using IBM SPSS software package version 27.0. (Armonk, NY: IBM Corp) Qualitative data were described using number and percent. The Kolmogorov-Smirnov test was used to verify the normality of distribution Quantitative data

were described using range (minimum and maximum), mean and standard deviation. Significance of the obtained results was judged at the 5% level (p $\leq$ 0.05). (17) The used tests were:

- **1- Student t-test independent sample:** for testing significance between parametric (≥30) quantitative variables in two studied groups.
- 2- Pearson's Chi-square test ( $\chi$ 2): for categorical variables when  $\leq$ 20% of cells with 5 value or less, to compare between different groups.

**3- Fisher's Exact test:** for testing significance between categorical variables when ≥20% of cells with 5 value or less.

#### Results

Table (3) shows distribution of the studied populations according to age and sex characteristics in relation to groups. In COVID-19 group, the age ranged from 18 to 79 years with mean value  $43.81 \pm 16.03$  years, while in non-COVID-19 group, the age ranged from 18 to 77 years with the mean of  $41.52 \pm 16.80$ . There was no statistical significant difference between

the two groups concerning age (t=0.935 and P=0.351).

In COVID-19 group; males were 58.9% and females were 41.1%. While in non-COVID-19 group males were 53.3% and females were 46.7%. There was no statistical significant difference between the two groups concerning sex ( $\chi$ 2=0.564 and P=0.453). Table (4) shows that the most affected age group was 30 to < 45 years by a

age group was 30 to < 45 years by a percentage of 34.4% followed by the age group less than 30 years by 31.1%, then group 45 to < 60 years by 22.2% and the least was 60 or more years by 12.2%.

Table (3): Age and sex profile of the 180 COVID-19 and non-COVID-19 patients.								
Age and Sex	COVID-19	COVID-19 n=90		/ID n=90	Analytic test	P value		
	n=90			11011-90	Analytic test	r value		
Age (years)								
Range	18	-79	18-77		t= 0.935	0.254		
Mean ± S.D.	43.81	43.81 ± 16.03		± 16.80		0.351		
Sex	N	%	N	%				
Male n=101	53	58.9	48	53.3	y² -0.564	0.453		
Female n=79	37	41.1	42	46.7	$\chi^2 = 0.564$ 0.45			
t= Student t test, χ2= Pearson Chi-Square test, S.D.= standard deviation.								
P value is considered significant if ≤0.05								

Table (4): Age and sex distribution of the 90 COVID-19 patients.							
Ago group (voors)	Sex (N=90	Sex (N=90)					
Age group (years)	Male	Females	Total	Percent			
Less than 30	16	12	28	31.1%			
30 to <45	20	11	31	34.4%			
45 to <60	11	9	20	22.2%			
60 or more	6	5	11	12.2%			
Total number in males and females	53	37	90				
Total percent in males and females	58.9%	41.1%	100%				

Table (5) shows that concerning the the 90 COVID-19 patients, azithromycin was significantly used by 82.2% of cases (P≤0.001), followed by amoxicillinclavulanic, ceftriaxone, ampicillin and doxycycline (42.2%, 12.2%, 7.8% and 7.8%

respectively). The least to be used was ciprofloxacin by 3.3% of cases.

While regarding antibiotic usage in non-COVID-19 group, metronidazols were used by 63.32% of cases, followed by macrolides (53.32%), penicillins (44.4%), and the least

used were  $3^{rd}$  generation cephalosporins (27.77%).

Table (6) shows the resistance profile of E. coli strains isolated from both COVID 19 and non-COVID 19 patients. E. coli strains from COVID-19 group were resistant to cefazolin, ampicillin, amoxicillin-clavulanic acid, cefepime, doxycyline and trimethoprim-sulfamethoxazol (91.1%, 61.1%, 38.9%, 35.6%, 32.2%, and 31.1% respectively). Nearly a similar pattern was obtained from E. coli strains from non -COVID-19 group for the same antibiotics (90%,73.3%, 52.2%, 40%, 40%, respectively).

Significantly higher resistance to ertapenem was observed in COVID group

versus non-COVID group (8.9% vs 0%) with P value equals 0.007. Significant higher resistance was observed in non-COVID group considering gentamicin (24.4% vs 7.8%, P value equals 0.004), ciprofloxacin (30% vs 15.6%, P value equals 0.032) and aztreonam (36.7% vs 21.1%, P value equals 0.032).

Table (7) illustrates that *E. coli* isolates from both COVID-19 and non-COVID-19 patients showed MDR profile (55.3% and 44.7% respectively) with a significantly higher percent of MDR among COVID -19 patients isolates (P=0.037).

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Table (5): Distribution of a	ntibiotics in	take by the	180 COVII	D-19 and non-0	COVID-19 patien	ts.
Antibiotic	COVID-19n=90		Non-CO	VID-19 n=90	Applytic tost	Dualica
	N	%	N	%	Analytic test	P value
Macrolides						
Azithromycin	74	82.22	8	8.88	χ²=97.57	≤0.001*
Clarithromycin	-	-	40	44.44	-	-
Penicillins				•		
Amoxicillin-Clavulanic	28	42.22	40	44.44	y²- 0 00	0.764
acid	38	42.22	40	44.44	$\chi^2 = 0.09$	0.764
Ampicillin	7	7.77	-	-	-	-
	3 <sup>rd</sup>	generation	cephalosp	oorins		
Ceftriaxone	11	12.22	18	20	χ²=2.014	0.155
Cefotaxime	-	-	7	7.77	-	-
Quinolones						
Ciprofloxacine	3	3.33	-	-	-	-
Tetracyclines						
Doxycycline	7	7.77	-	-	-	-
Metronidazoles						
Metronidazole	-	-	57	63.32	-	-
				•		

χ2= Pearson Chi-Square test.

P value is considered significant if ≤0.05.

P value is considered highly significant if ≤0.005

Some patients take more than one antibiotic

• •		e profile of E-coli st	trains isolate	ed from th	e 180 stoo	I samples of	the 180				
COVID-19 and	COVID-19 and non-COVID-19 studied patients.  Disc Resistance profile										
Mode of action	Antibiotic class	Antibiotic name	Disc content	COVID-19	e profile Non- COVID-19	Test	P value				
		Ampicillin (AMP)	10 μg	55 (61.1%)	66 (73.3%)	χ2=3.051	0.112				
	Penicillins	Amoxicillin clavulanic (AMC)	20/10 μg	35 (38.9%)	47 (52.2%)	Fissure Exact = 3.921	0.099				
		Ceftriaxone (CRO)	30 µg	26 (28.9%)	37 (41.1%)	Fissure Exact =3.700	0.118				
Inhibition of cell wall synthesis	Cephalosporins	Cefazolin (CFZ)	30 µg	82 (91.1%)	81 (90%)	Fissure Exact = 2.474	0.591				
		Cefepime (FEP)	30 µg	32 (35.6%)	36 (40%)	χ2=0.378	0.645				
	Carbapenems	Ertapenem (ETP)	10 μg	8 (8.9%)	o (o%)	Fissure Exact =11.463	0.007*				
	Monobactams	Aztreonam (AZT)	30 µg	19 (21.1%)	33 (36.7%)	χ2=5.300	0.032*				
Inhibition of protein	Aminoglycosides	Gentamicin (CN)	10 µg	7 (7.8%)	22 (24.4%)	χ2=9.249	0.004*				
synthesis	Tetracyclines	Doxycycline (DOX)	30 µg	29 (32.2%)	36 (40%)	χ2=1.180	0.352				
Inhibition of cell DNA replication	Quinolones	Ciprofloxacin (CIP)	5 μg	14 (15.6%)	27 (30%)	χ2=5.338	0.032*				
Inhibition of folic acid metabolism	Folate pathway Sulfamethoxazole		25/23.75 μg	28 (31.1%)	29 (32.2%)	Fissure Exact = 0.995	0.873				
χ2= Pearson	χ₂= Pearson Chi-Square test. P value is considered significant if ≤0.05.										

P value is considered significant if ≤0.05.

χ2= Pearson Chi-Square test.
P value is considered highly significant if ≤0.005

Table (7): Multidrug resistance profile of <i>E. coli</i> isolated from the 180 COVID-19 and non-COVID-19 studied patients								
	COVID-19		Non-COVI	D-10 n-00				
Drug resistance	n=90		Non-COVID-19 n=90		Analytic test	P value		
	N	%	N	%				
MDR	68	55.3%	55	44.7%	y²- 4 220	0.027*		
Not MDR	22	38.6%	35	61.4%	χ²=4.339	0.037*		
χ2= Pearson Chi-Square test. P value is considered significant if ≤0.05.								
P value is considered highly significant if ≤0.005								

#### Discussion

The global community continues to grapple with the aftermath of the COVID-19 pandemic, which originated in Wuhan, Hubei, China, at a seafood market. From its initial detection in 2019 to October 2022, the pandemic tragically resulted in nearly 6 million deaths worldwide. Additionally, a staggering 755,703,002 confirmed cases of COVID-19 were reported during this period.<sup>(18)</sup>

In our study, the patients' age within the COVID-19 group varied between 18 and 79 years. The most affected age group was the group from 30 up to < 45 years (34.4%), followed by the age group from 18 to < 30 years (31.1%) as shown in table (4). These findings are in agreement with the Centers for Disease Control and Prevention (CDC) report in September 2020. This report stated that during June–August 2020, the COVID-19 prevalence was greater in young adults (20–29 years) accounting for more than 20% of all confirmed COVID-19 cases. (19)

Globally, numerous epidemiological reports have established that the change in age distribution occurred in a time-dependent pattern. As the pandemic first began, the majority of COVID-19 patients were adults. Then, the virus was simultaneously spreading to younger people as the COVID-19 variant was progressing. (20)

Ahmad S <sup>(21)</sup> have stated that the observed differences within the age profile between the different patient's groups were confounded by several factors as follows: the degree and target of tested population, ethnic variations from one place to the other, and age variations within the general population.

Several concerned reports of the COVID-19 infection have abandoned the possible sexrelated influences although sex differences were described in both diagnosis and testing of COVID-19. (22)

In agreement with Sama et al. (23), our study findings showed that male patients were more affected by COVID-19 than females (58.9% and 41.1% respectively) as illustrated in tables (3 and 4). It has been observed that men, globally, have a relatively greater consumption rate of both smoking and Moreover, men are alcohol. susceptible to high-risk behaviors and certain occupational hazards such as transportation, construction, manufacturing delivery and food processing than women. This is further elevating men's likelihoods to contract the COVID-19. (24)

On the other hand, several reports have confirmed that women display greater adherence to the COVID-19 linked controlling measures, such as face maskwearing, social distancing, hand washing, and restrictions on movement. These COVID-19 proper behaviors aid in protecting individuals from COVID-19 contracting. (25)

In the present study, azithromycin was significantly administered to the COVID-19 patients (82.2%) compared to the non-COVID-19 patients (8.88%) with P  $\leq$  0.001 as shown in table (5). It is worth mentioning that in COVID-19 infection, azithromycin is approved by the Egyptian Ministry of Health and Population (MOHP) to be among the treatment protocol modalities for the pandemic. (26)

In their work, Kamara et al. (27) have reported similar results where azithromycin was the most commonly supplemented antibiotic (34.8%). In contrast, Saleem et al. (28) reported that

among COVID-19 patients, the top 3 most frequently prescribed antimicrobials were ceftriaxone (26.6%), metronidazole (9.7%) and vancomycin (7.9%). Their study was performed in Pakistani hospitals where ceftriaxone was usually used as intravenous injection.

Considering using azithromycin in COVID patients' treatment, Butler et al. (29) have stated that azithromycin must not be routinely utilized to manage COVID-19 in the community, in lack of further indications. Their findings have had vital stewardship implications for antibiotic throughout this pandemic. It indicated that the inadequate antibiotics usage could lead to augmented resistance for the antibiotic. As a severe worldwide threat, AMR is regarded as growing health care concern. Unfortunately, this matter has not received sufficient attention throughout the last COVID-19 pandemic. During the COVID-19 pandemic, researchers have discovered a noteworthy decline in AMR surveillance. This might have restricted the capability to provide data on AMR definite alterations and it raised the likelihood of having a silent AMR pandemic. (30)

In this study, *E. coli* was isolated from stool samples for examining its antibiotic resistance profile. *E. coli* is one of the most globally investigated bacteria. Debatably, it is the best comprehended of the whole model microorganisms. (31)

The extrinsic (acquired) resistance along with constantly rising *E. coli* resistance to antibiotics is presently regarded as a major public health issue worldwide. One of the main factors contributing to the current antibacterial resistance dilemma is the fast evolution of bacteria under selective antibiotic pressure. Since a key factor contributing to the rise in MDR strains of

bacteria is the ongoing interaction between any specific antibiotic and the bacterium. Regrettably, the inappropriate prescription and overuse of these medications are 2 large contributors to such problem. (10)

Our study showed (in table 6) that AMR of *E.coli* was detected in both COVID-19 and non-COVID-19 groups with no significant difference except for ertapenem which was observed in COVID group vs non-COVID group by (8.9% vs 0%, P = 0.007) and on the other hand, significant higher resistance was observed in non-COVID group considering gentamicin (24.4% vs 7.8%, P = 0.004), ciprofloxacin (30% vs 15.6%, P = 0.032) and aztreonam (36.7% vs 21.1%, P = 0.032).

These results are in agreement with Kishk et al. (32). They have reported that on comparison between antibiotic susceptibility of gram-negative organisms within the non-COVID-19 and during the COVID-19 era, there was not significant differences between the studied groups regarding ceftriaxone, cefepime, ampicillin and trimethoprim.

Even though quinolones are a well-known class of broad-spectrum antibiotics, the current study stated resistance rate was rather high (30%). There are growing concerns about the widespread overuse of these broad-spectrum antibiotics in developing nations, which could result in a notable rise in the rate of resistance. Bolon et al., for example, have documented a constant, stepwise rise in *E. coli* resistance to ciprofloxacin. (33)

In our study, carbapenems continued to be the most effective medication against *E. coli*. This is consistent with the findings of Garcia et al. <sup>(34)</sup>, who reported that carbapenem resistance was not observed. Carbapenem resistance deteriorates with

time, posing threats to the infected individuals and serving as some MDR isolates' sole antibiotic option. More than 50% of the reported isolates of *E. coli* to the European Centre for Disease Prevention and Control (ECDC) in 2018 demonstrated resistance to at least one antimicrobial group under monitoring, and MDR was common. (35)

In our research, the MDR *E. coli* isolates were considerably greater in the COVID-19 individuals (55.3%) in comparison to the non-COVID-19 individuals (44.7%) (P =0.037), though both findings were high (as shown in table 7). MDR is a reflection of the widespread use of antibiotics, and its great prevalence is due to the organism's capacity to acquire resistance genes. (36)

These results are in line with Bogossian et al. (37) who evaluated the acquisition of MDR bacteria in patients with COVID-19. Their MDR percentage was 54%, which is extremely close to the MDR percentage in our study's COVID-19 patients (55.3%).

These findings differed from those of Alali et al. <sup>(38)</sup>, who found that the MDR prevalence of *E. coli* isolated from COVID-19 medical wards was 37.4%, with no noticeable difference from non-COVID-19 wards.

Overusing antibiotics is considered the main cause of antibiotic resistance, which might affect antibiotic stewardship and hence worsen the AMR.

#### Conclusion

E. coli isolated from stool samples of COVID-19 patients have shown a significantly higher MDR profile than non-COVID-19 patients. There was a substantial difference in azithromycin usage between COVID-19 and non-COVID-19 patients, with COVID-19 patients exhibiting higher usage rates. A detected high resistance to the broad-spectrum quinolones was observed

in both COVID-19 and non-COVID-19 patients, while the most efficient drugs against *E. coli* were the Carbapenems in both groups.

#### Recommendations

Antibiotic use must be restricted without the clinician appropriate indication particularly those antibiotics used as a last resort. MDR organisms in the community necessitate the importance of adopting educational programs to raise awareness about the hazards of antibiotic abuse including bacterial resistance.

#### References

- 1. World Health Organization [WHO]. Antimicrobial resistance. Geneva: WHO; 2021.
- Centers for Disease Control and Prevention [CDC]. 2019 AR Threats Report. Atlanta: CDC; 2019.
- 3. Lucien MAB, Canarie MF, Kilgore PE, et al. Antibiotics and antimicrobial resistance in the COVID-19 era: Perspective from resource-limited settings. Int J Infect Dis. 2021;104:2504.
- 4. Furlan L., Caramelli B. The Regrettable Story of the "COVID Kit" and the "Early Treatment of COVID-19" in Brazil. Lancet Reg. Health—Am. 2021;4:100089. doi: 10.1016/j.lana.2021.100089.
- 5. Youngs J, Wyncoll D, Hopkins P, Arnold A, Ball J, Bicanic T. Improving antibiotic stewardship in COVID-19: Bacterial co-infection is less common than with influenza. J Infect. 2020;81(3):e55-7.
- 6. Olesen SW, Barnett ML, MacFadden DR, et al. The distribution of antibiotic use and its association with antibiotic resistance. Elife. 2018;7:e39435.
- Sommer MOA, Dantas G, Church GM. Functional characterization of the antibiotic resistance reservoir in the human microflora. Science. 2009;325(5944):1128-31.

- 8. Horesh G, Blackwell GA, Tonkin-Hill G, Corander J, Heinz E, Thomson NR. A comprehensive and high-quality collection of Escherichia coli genomes and their genes. Microb Genom. 2021;7(2):000499.
- Van Seventer JM, Hochberg NS. Principles of infectious diseases: transmission, diagnosis, prevention, and control. Int Encycl Public Health. 2017;2017:22-39.
- 10. Kolár M, Urbánek K, Látal T. Antibiotic selective pressure and development of bacterial resistance. Int J Antimicrob Agents. 2001;17(5):357-63.
- 11. Klein EY, Van Boeckel TP, Martinez EM, et al. Global increase and geographic convergence in antibiotic consumption between 2000 and 2015. Proc Natl Acad Sci U S A. 2018;115(15):E3463-70.
- 12. Nordmann P, Poirel L. Epidemiology and Diagnostics of Carbapenem Resistance in Gram-negative Bacteria. Clin Infect Dis. 2019;69(Suppl 7):S521-8.
- 13. Shakya P, Barrett P, Diwan V, et al. Antibiotic resistance among Escherichia coli isolates from stool samples of children aged 3 to 14 years from Ujjain, India. BMC Infect Dis. 2013;13(1):477.
- 14. Alzaher MZ, Almugahwi AA, Almulla AA, Almeer HH, Alshammasi MM, El-Badry AA. Diagnostic yield of stool culture and probable predictive factors, a single-center experience. Acta Bio Medica Atenei Parm. 2022;93(6)
- 15. Tille PM. Bailey and Scott's Diagnostic Microbiology. 13th ed. St. Loius: Mosby Company; 2013.
- 16. Clinical and Laboratory Standards Institute [CLSI]. Performance standards for antimicrobial susceptibility testing. 32nd ed. Wayne: CLSI; 2022.
- 17. Kirkpatrick LA, Feeney BC. A simple guide to IBM SPSS statistics for version 27.0. Released 2019, modification 18 April 2022.
- 18. World Health Organization [WHO]. COVID-19 Dashboard. Geneva, Switzerland: WHO; 2020.
- 19. Furuse Y, Sando E, Tsuchiya N, et al. Clusters of coronavirus disease in

- communities, Japan, January–April 2020. Emerg Infect Dis. 2020;26:2176–9.
- 20. Martin B, DeWitt PE, Russell S, et al. Characteristics, outcomes, and severity risk factors associated with SARS-CoV-2 infection among children in the US national COVID cohort collaborative. JAMA Netw Open. 2022;5(2):e2143151.
- 21. Ahmad S. Potential of age distribution profiles for the prediction of COVID-19 infection origin in a patient group. Inform Med Unlocked. 2020;20:100364.
- 22. Ballering AV, Oertelt-Prigione S, Olde Hartman TC, Rosmalen JGM. Sex and Gender-Related Differences in COVID-19 Diagnoses and SARS-CoV-2 Testing Practices During the First Wave of the Pandemic: The Dutch Lifelines COVID-19 Cohort Study. J Womens Health (Larchmt). 2021;30(12):1686-92.
- 23. Sama IE, Ravera A, Santema BT, et al. Circulating plasma concentrations of angiotensin-converting enzyme 2 in men and women with heart failure and effects of renin-angiotensin-aldosterone inhibitors. Eur Heart J. 2020;41(19):1810-7.
- 24. Takahashi T, Ellingson MK, Wong P, et al. Sex differences in immune responses that underlie COVID-19 disease outcomes. Nature. 2020;588(7837):315-20.
- 25.Griffith DM, Sharma G, Holliday CS, et al. Men and COVID-19: A Biopsychosocial Approach to Understanding Sex Differences in Mortality and Recommendations for Practice and Policy Interventions. Prev Chronic Dis. 2020;17:E63.
- 26. Sultana J, Cutroneo PM, Crisafulli S, Puglisi G, Caramori G, Trifirò G. Azithromycin in COVID-19 Patients: Pharmacological Mechanism, Clinical Evidence and Prescribing Guidelines. Drug Saf. 2020;43(8):691-8.
- 27.Kamara IF, Kumar AM, Maruta A, et al. Antibiotic Use in Suspected and Confirmed COVID-19 Patients Admitted to Health Facilities in Sierra Leone in 2020-2021: Practice Does Not Follow Policy. Int J Environ Res Public Health. 2022;19(7):4005.

- 28. Saleem Z, Haseeb A, Godman B, et al. Point Prevalence Survey of Antimicrobial Use during the COVID-19 Pandemic among Different Hospitals in Pakistan: Findings and Implications. Antibiotics. 2022;12(1):70.
- 29. Butler CC, Dorward J, Yu LM, et al. Azithromycin for community treatment of suspected COVID-19 in people at increased risk of an adverse clinical course in the UK (PRINCIPLE): a randomised, controlled, open-label, adaptive platform trial. Lancet. 2021;397(10279):1063-74.
- 30. Tomczyk S, Taylor A, Brown A,, et al. Impact of the COVID-19 pandemic on the surveillance, prevention and control of antimicrobial resistance: a global survey. J Antimicrob Chemother. 2021;76(11):3045-58.
- 31. Clancy CJ, Nguyen MH. Coronavirus Disease 2019, superinfections, and antimicrobial development: What can we expect? Clin Infect Dis. 2020;71(10):2736-43.
- 32.Kishk R, Abu Bakr NM, Anani M, et al. Pattern of antimicrobial resistance in the pre and during COVID-19 era: An observational study. Microbes Infect Dis. 2023;4(4):1100-13.
- 33.Bolon MK, Wright SB, Gold HS, Carmeli Y. The magnitude of the association between fluoroquinolone use and quinolone-resistant Escherichia coli and Klebsiella pneumoniae may be lower than previously reported. Antimicrob Agents Chemother. 2004;48(6):1934-40.
- 34. Garcia PG, Silva VL, Diniz CG. Occurrence and antimicrobial drug susceptibility patterns of commensal and diarrheagenic Escherichia coli in fecal microbiota from children with and without acute diarrhea. J Microbiol. 2011;49(1):46-52.
- 35. European Centre for Disease Prevention and Control [ECDC]. Surveillance of antimicrobial resistance in Europe 2018. Stockholm: ECDC; 2019.
- 36. Exner M, Bhattacharya S, Christiansen B, et al. Antibiotic resistance: What is so special about multidrug-resistant Gram-

- negative bacteria? GMS Hyg Infect Control. 2017;12:Doco5.
- 37. Bogossian EG, Taccone FS, Izzi A, et al. The acquisition of multidrug-resistant bacteria in patients admitted to COVID-19 intensive care units: A monocentric retrospective case control study. Microorganisms. 2020;8(11):1821.
- 38. Alali WQ, Abdo NM, AlFouzan W, Dhar R. Antimicrobial resistance pattern in clinical Escherichia coli and Pseudomonas aeruginosa isolates obtained from a secondary-care hospital prior to and during the COVID-19 pandemic in Kuwait. Germs. 2022;12(3):372-83.